

Ecography

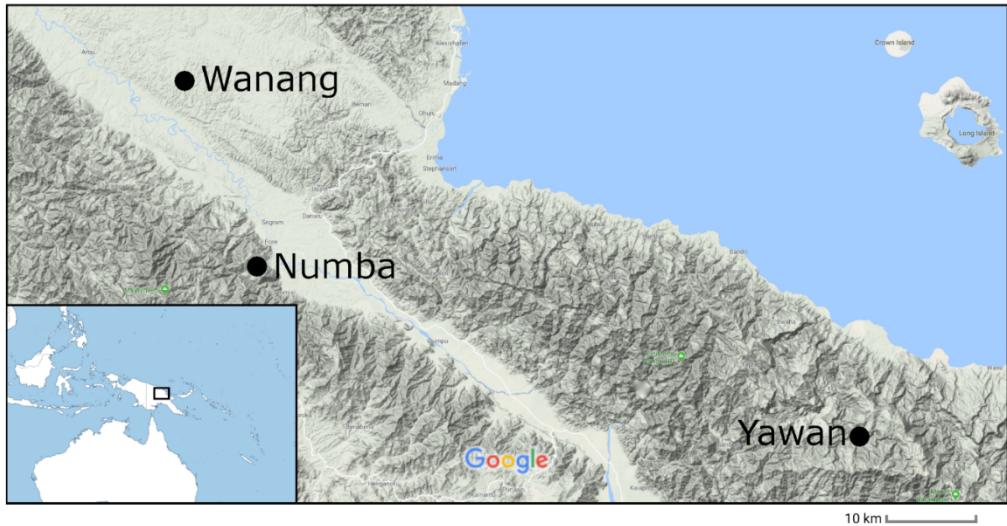
**ECOG-04730**

Plowman, N. S., Mottl, O., Novotny, V., Idigel, C., Philip, F. J., Rimandai, M. and Klimes, P. 2019. Nest microhabitats and tree size mediate shifts in ant community structure across elevation in tropical rainforest canopies. – *Ecography* doi: 10.1111/ecog.04730

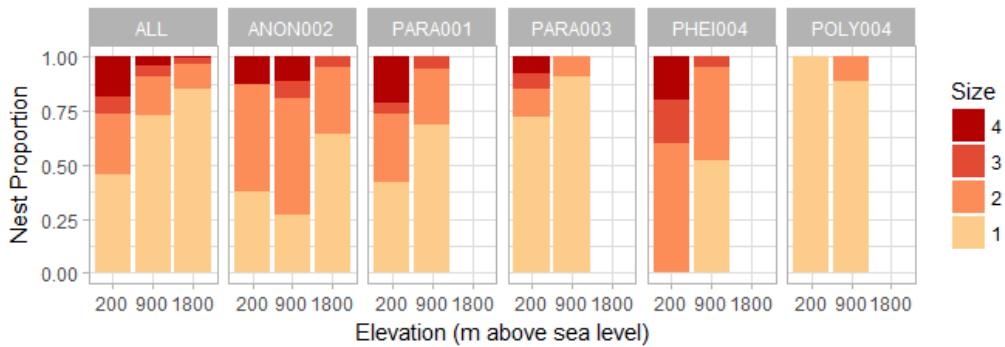
**Supplementary material**

## Appendix A1 - Supplementary Figures & Tables

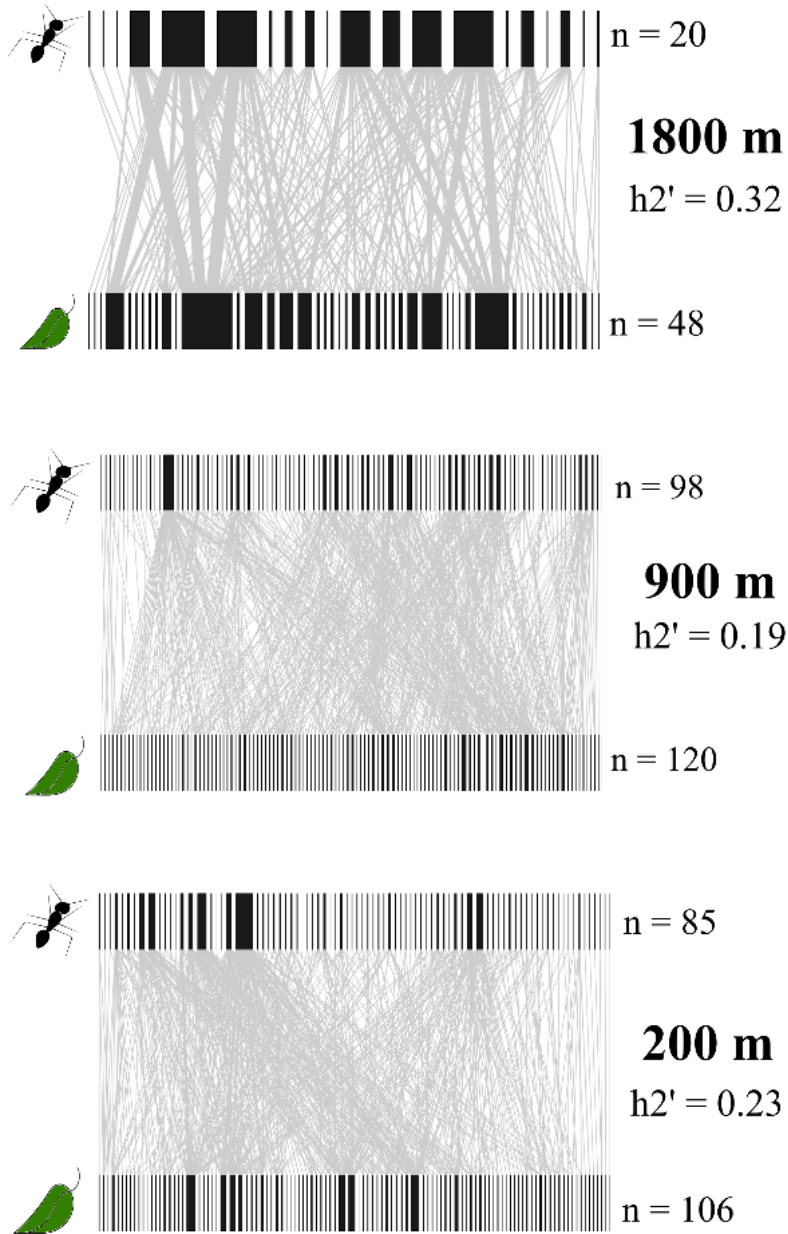
### Supplementary Figures



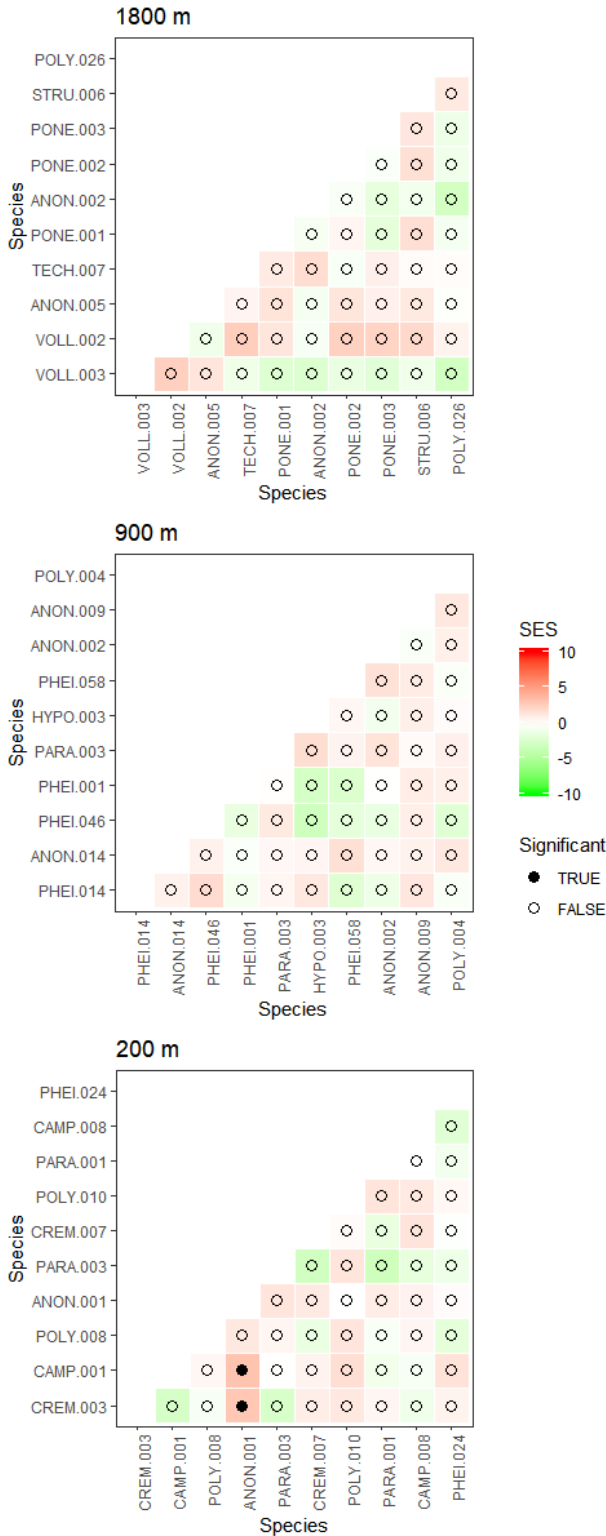
**Figure A1.** Map indicating location of study sites in north-eastern Papua New Guinea. We sampled two lowland forest plots near Wanang village (~200 m.a.s.l; 05°140S 145°110E), one mid-elevation forest plot near Numba village, Bismarck Range (~900 m.a.s.l; 5°43'18"S, 145°16'2"E), and two high elevation forest plots near Yawan village, Saruwaged Range (~1800 m a.s.l; 6°09'46"S146°50'22" E). Image adapted from Map data © 2018 Google.



**Figure A2.** Change in nest size distribution of tree-nesting ants over elevation. Nest size categories are based on the estimated number of workers per nest (1:  $\leq 100$ , 2:  $\sim 100-500$ , 3:  $\sim 500-1000$ , 4:  $> 1000$ ). Nest size significantly differed between three elevations ( $\chi^2=297.64$ ,  $df= 6$ ,  $P < 0.001$ ,  $n=2143$ ) with a decreasing proportion of large nests (cat. 2-4) and an increase in small nests (cat. 1). A similar pattern was found for the most common species present at more than one elevation: *Anonychomyrma scrutator* (ANON002,  $n=79$ ), *Paraparatrechina pallida* (PARA001,  $n=57$ ), *Paraparatrechina minutula* (PARA003,  $n=74$ ), *Pheidole hospes* (PHEI004,  $n=31$ ) and *Polyrhachis debilis* (POLY004,  $n=40$ ).



**Figure A3.** Bipartite networks with elevation between ants and trees hosting their nests, with  $h2'$  network specialisation index. Upper bars represent ant species, lower bars represent the tree species they are found nesting in. All elevations have low network specialisation (0 being completely unspecialised, 1 being fully specialised), indicating that ant – tree species specificity was very low in all three sites.



**Figure A4.** Results of co-occurrence model for pairs of the ten most common tree nesting ant species at low, mid and high elevation. Positive standardised effect size values in red (SES) indicate negative association (i.e. species co-occur in trees less often than expected by chance), and negative SES indicates positive associations (i.e. species co-occur more often than expected by chance). The only significantly non-random associations ( $P < 0.001$ ) occurred at low elevation and are outlined in black – *Anonychomyrma cf. scrutator* (ANON001) was negatively associated with *Crematogaster polita* (CREM003) and *Colobopsis vitrea* (CAMP001). ANON001 and CREM003 were the most common dominant ant species at low elevations. See Table A1 for species codes, taxonomic names, and abundance at each elevation.

## Supplementary Tables

**Table A1.** Full list of arboreal nesting ant species (ordered alphabetically), and their nest abundance at each elevation. Mounted voucher specimens and additional material in ethanol are available for each Species code at the Institute of Entomology, Biology Centre of the Czech Academy of Sciences, Ceske Budejovice. Accession numbers for each site are as follows: Wanang – HP0001 to HP0732; Numba – NA0012 to NA0242; Yawan – YA0001 to YA0097.

Species	Subfamily	Genus	Species code	200 m	900 m	1800 m
<i>Acropyga ambigua</i> Emery, 1922	Formicinae	<i>Acropyga</i>	ACRO 001		1	
<i>Aenictus nesiotis</i> Wheeler & Chapman 1930	Dorylinae	<i>Aenictus</i>	AENI 003		1	
<i>Anochetus cato</i> Forel, 1901	Ponerinae	<i>Anochetus</i>	ANOC 001		8	
<i>Anochetus</i> sp. 3	Ponerinae	<i>Anochetus</i>	ANOC 003		1	
<i>Anonychomyrma dimorpha</i> (Viehmeyer, 1912)	Dolichoderinae	<i>Anonychomyrma</i>	ANON 003			16
<i>Anonychomyrma minuta</i> (Donisthorpe, 1943)	Dolichoderinae	<i>Anonychomyrma</i>	ANON 002	8	24	45
<i>Anonychomyrma</i> cf. <i>scrutator</i> (Smith F., 1859)	Dolichoderinae	<i>Anonychomyrma</i>	ANON 001	50	5	
<i>Anonychomyrma</i> sp. 5	Dolichoderinae	<i>Anonychomyrma</i>	ANON 005			66
<i>Anonychomyrma</i> sp. 9	Dolichoderinae	<i>Anonychomyrma</i>	ANON 009		26	
<i>Anonychomyrma</i> sp. 12	Dolichoderinae	<i>Anonychomyrma</i>	ANON 012		7	
<i>Anonychomyrma</i> sp. 14	Dolichoderinae	<i>Anonychomyrma</i>	ANON 014		51	
<i>Anonychomyrma</i> sp. 15 aff. <i>dimorpha</i> (Viehmeyer, 1912)	Dolichoderinae	<i>Anonychomyrma</i>	ANON 015		22	
<i>Brachyponera croceicornis</i> (Emery, 1900)	Ponerinae	<i>Brachyponera</i>	PACH 006	1	3	
<i>Calomyrmex laevisimus</i> (Smith, 1859)	Formicinae	<i>Calomyrmex</i>	CALO 001		3	
<i>Camponotus</i> ( <i>Forelophilus</i> ) sp. 1	Formicinae	<i>Camponotus</i>	FORE 001		2	
<i>Camponotus anezkae</i> Klimes & McArthur, 2014	Formicinae	<i>Camponotus</i>	CAMP 022	1		
<i>Camponotus dorycus confusus</i> Emery, 1887	Formicinae	<i>Camponotus</i>	CAMP 016	7	1	

<i>Camponotus cf. flavocassisi</i> Donisthorpe, 1941	Formicinae	<i>Camponotus</i>	CAMP 033	3	
<i>Camponotus hastifer</i> Emery, 1911	Formicinae	<i>Camponotus</i>	CAMP 038	4	
<i>Camponotus mussolinii</i> Donisthorpe, 1936	Formicinae	<i>Camponotus</i>	CAMP 028	4	
<i>Camponotus cf. variegatus</i> (Smith, F., 1858)	Formicinae	<i>Camponotus</i>	CAMP 018	2	12
<i>Camponotus triangulatus</i> Klimes & McArthur, 2014	Formicinae	<i>Camponotus</i>	CAMP 019	1	
<i>Camponotus wanangus</i> Klimes & McArthur, 2014	Formicinae	<i>Camponotus</i>	CAMP 003	3	2
<i>Camponotus sp. 7 aff. trajanus</i> Forel, 1912	Formicinae	<i>Camponotus</i>	CAMP 007	12	
<i>Camponotus sp. 20 aff. janeti</i> Forel, 1895	Formicinae	<i>Camponotus</i>	CAMP 020	1	
<i>Camponotus sp. 32</i>	Formicinae	<i>Camponotus</i>	CAMP 032		1
<i>Cardiocondyla sp. 2 aff. thoracina</i> (Smith, 1859)	Myrmicinae	<i>Cardiocondyla</i>	CARD 002		1
<i>Carebara atoma</i> Emery, 1900	Myrmicinae	<i>Carebara</i>	CARE 002	1	
<i>Carebara crassiuscula</i> (Emery, 1900)	Myrmicinae	<i>Carebara</i>	CARE 006		1
<i>Carebara sp. 1</i>	Myrmicinae	<i>Carebara</i>	CARE 001		3
<i>Carebara sp. 3</i>	Myrmicinae	<i>Carebara</i>	CARE 003		1
<i>Cerapachys cf. flavaclavatus</i> Donisthorpe, 1938	Dorylinae	<i>Cerapachys</i>	CERA 001	1	2
<i>Cerapachys opacus</i> Emery, 1901	Dorylinae	<i>Cerapachys</i>	CERA 004		1
<i>Cerapachys sp. 5</i>	Dorylinae	<i>Cerapachys</i>	CERA 005		1
<i>Colobopsis aruensis</i> Karavaiev, 1933	Formicinae	<i>Colobopsis</i>	CAMP 004	3	
<i>Colobopsis conithorax</i> Emery, 1914	Formicinae	<i>Colobopsis</i>	CAMP 006	10	
<i>Colobopsis cf. macrocephala</i> (Erichson, 1842)	Formicinae	<i>Colobopsis</i>	CAMP 010	5	6
<i>Colobopsis aff. polynesica</i> (Emery, 1896)	Formicinae	<i>Colobopsis</i>	CAMP 017	1	
<i>Colobopsis quadriceps</i> (Smith F., 1859)	Formicinae	<i>Colobopsis</i>	CAMP 013	1	3
<i>Colobopsis rotunda</i> Klimes & McArthur, 2014	Formicinae	<i>Colobopsis</i>	CAMP 014	2	
<i>Colobopsis vitrea</i> (Smith F., 1860)	Formicinae	<i>Colobopsis</i>	CAMP 001	75	6

<i>Colobopsis</i> sp. 5 aff. <i>conithorax</i> Emery, 1914	Formicinae	<i>Colobopsis</i>	CAMP 005	5	
<i>Colobopsis</i> sp. 8 aff. <i>sanguinifrons</i> Viehmeyer, 1925	Formicinae	<i>Colobopsis</i>	CAMP 008	27	
<i>Colobopsis</i> sp. 24 aff. <i>aruensis</i>	Formicinae	<i>Colobopsis</i>	CAMP 024		4
<i>Colobopsis</i> sp. 37 aff. <i>sanguinifrons</i> Viehmeyer, 1925	Formicinae	<i>Colobopsis</i>	CAMP 037	2	
<i>Crematogaster elysii</i> Mann, 1919	Myrmicinae	<i>Crematogaster</i>	CREM 002	18	3
<i>Crematogaster emeryi</i> Forel, 1907	Myrmicinae	<i>Crematogaster</i>	CREM 010		1
<i>Crematogaster flavitarsis</i> Emery, 1900	Myrmicinae	<i>Crematogaster</i>	CREM 005		1
<i>Crematogaster polita</i> Smith F., 1865	Myrmicinae	<i>Crematogaster</i>	CREM 003	146	
<i>Crematogaster</i> cf. <i>pythia</i> Forel, 1915	Myrmicinae	<i>Crematogaster</i>	CREM 004	2	
<i>Crematogaster tarsata</i> Smith, F. 1865	Myrmicinae	<i>Crematogaster</i>	CREM 013		6
<i>Crematogaster</i> sp. 6	Myrmicinae	<i>Crematogaster</i>	CREM 006	6	
<i>Crematogaster</i> sp. 7 aff. <i>fritzzi</i> Emery, 1901	Myrmicinae	<i>Crematogaster</i>	CREM 007	39	2
<i>Crematogaster</i> sp. 11 aff. <i>fritzzi</i> Emery, 1901	Myrmicinae	<i>Crematogaster</i>	CREM 011	3	
<i>Crematogaster</i> sp. 15 aff. <i>flavicornis</i> Emery, 1897	Myrmicinae	<i>Crematogaster</i>	CREM 015		3
<i>Diacamma rugosum</i> (Le Guillou, 1842)	Ponerinae	<i>Diacamma</i>	DIAC 001	13	
<i>Echinopla</i> sp. 2	Formicinae	<i>Echinopla</i>	ECHI 002	1	
<i>Echinopla</i> sp. 3	Formicinae	<i>Echinopla</i>	ECHI 003		2
<i>Hypoponera</i> cf. <i>confinis</i> Roger, 1860	Ponerinae	<i>Hypoponera</i>	HYPO 002	2	15
<i>Hypoponera sabrone</i> Donisthorpe, 1941	Ponerinae	<i>Hypoponera</i>	HYPO 003	1	31
<i>Hypoponera</i> sp. 4	Ponerinae	<i>Hypoponera</i>	HYPO 004		9
<i>Leptomyrmex flavitarsus</i> (Smith, F., 1859)	Dolichoderinae	<i>Leptomyrmex</i>	LEPM 002		1
<i>Lordomyrma</i> sp. 7	Myrmicinae	<i>Lordomyrma</i>	LORD 007		1
<i>Lordomyrma</i> sp. 10	Myrmicinae	<i>Lordomyrma</i>	LORD 010	29	
<i>Monomorium intrudens</i> Smith F., 1894	Myrmicinae	<i>Monomorium</i>	MONO 002	2	



<i>Monomorium pharaonis</i> (Linnaeus 1758)	Myrmicinae	<i>Monomorium</i>	MONO 004	1	
<i>Monomorium</i> sp. 3	Myrmicinae	<i>Monomorium</i>	MONO 003	3	
<i>Myopias</i> cf. <i>delta</i> Willey & Brown, 1983	Ponerinae	<i>Myopias</i>	MYOP 003		1
<i>Myrmecina mandibularis</i> Viehmeyer, 1914	Myrmicinae	<i>Myrmecina</i>	MYRM 003		5
<i>Nylanderia</i> aff. <i>vaga</i> (Forel, 1901)	Formicinae	<i>Nylanderia</i>	PARA 005	7	
<i>Nylanderia nuggeti</i> Donisthorpe, 1941	Formicinae	<i>Nylanderia</i>	PARA 007	2	13
<i>Odontomachus tyrannicus</i> Smith, 1859	Ponerinae	<i>Odontomachus</i>	ODON 005		5
<i>Oecophylla smaragdina</i> (Fabricius, 1775)	Formicinae	<i>Oecophylla</i>	OECO 001	2	12
<i>Paraparatrechina minutula</i> (Forel, 1901)	Formicinae	<i>Paraparatrechina</i>	PARA 003	40	34
<i>Paraparatrechina pallida</i> Donisthorpe, 1947	Formicinae	<i>Paraparatrechina</i>	PARA 001	38	19
<i>Paraparatrechina</i> sp. 2	Formicinae	<i>Paraparatrechina</i>	PARA 002	4	
<i>Paraparatrechina</i> sp. 6	Formicinae	<i>Paraparatrechina</i>	PARA 006	1	4
<i>Paraparatrechina</i> sp. 13	Formicinae	<i>Paraparatrechina</i>	PARA 013		4
<i>Paraparatrechina</i> sp. 14	Formicinae	<i>Paraparatrechina</i>	PARA 014		1
<i>Pheidole amber</i> Donisthorpe, 1941	Myrmicinae	<i>Pheidole</i>	PHEI 046		45
<i>Pheidole fuscula</i> Emery, 1900	Myrmicinae	<i>Pheidole</i>	PHEI 003		17
<i>Pheidole hospes</i> Smith, F. 1865	Myrmicinae	<i>Pheidole</i>	PHEI 004	10	21
<i>Pheidole sexspinosa biroi</i> Emery, 1900	Myrmicinae	<i>Pheidole</i>	PHEI 030		6
<i>Pheidole</i> sp. 1	Myrmicinae	<i>Pheidole</i>	PHEI 001		35
<i>Pheidole</i> sp. 2 aff. <i>sexspinosa biroi</i> Emery, 1900	Myrmicinae	<i>Pheidole</i>	PHEI 002	1	
<i>Pheidole</i> sp. 7 aff. <i>gambogia</i> Donisthorpe, 1948	Myrmicinae	<i>Pheidole</i>	PHEI 007	10	4
<i>Pheidole</i> sp. 14 aff. <i>gambogia</i> Donisthorpe, 1948	Myrmicinae	<i>Pheidole</i>	PHEI 014	5	91
<i>Pheidole</i> sp. 24 aff. <i>amber</i> Donisthorpe, 1941	Myrmicinae	<i>Pheidole</i>	PHEI 024	23	
<i>Pheidole</i> sp. 25 aff. <i>sexspinosa biroi</i> Emery, 1900	Myrmicinae	<i>Pheidole</i>	PHEI 025	2	

<i>Pheidole</i> sp. 26	Myrmicinae	<i>Pheidole</i>	PHEI 026	1		
<i>Pheidole</i> sp. 38	Myrmicinae	<i>Pheidole</i>	PHEI 038		2	
<i>Pheidole</i> sp. 48	Myrmicinae	<i>Pheidole</i>	PHEI 048			6
<i>Pheidole</i> sp. 49	Myrmicinae	<i>Pheidole</i>	PHEI 049			1
<i>Philidris</i> cf. <i>cordata</i> (Smith F., 1859)	Dolichoderinae	<i>Philidris</i>	PHIL 001	8	1	
<i>Philidris</i> sp. 2	Dolichoderinae	<i>Philidris</i>	PHIL 002	7		
<i>Philidris</i> sp. 3	Dolichoderinae	<i>Philidris</i>	PHIL 003	2		
<i>Plagiolepis</i> sp. 2	Formicinae	<i>Plagiolepis</i>	PLAG 002		8	
<i>Platythyrea parallela</i> (Smith, F., 1859)	Ponerinae	<i>Platythyrea</i>	PLAT 001		1	
<i>Podomyrma basalis</i> Smith F., 1859	Myrmicinae	<i>Podomyrma</i>	PODO 004		2	
<i>Podomyrma keysseri</i> Viehmeyer, 1914	Myrmicinae	<i>Podomyrma</i>	PODO 005			3
<i>Podomyrma</i> sp. 2 aff. <i>basalis</i> Smith F., 1859	Myrmicinae	<i>Podomyrma</i>	PODO 002	4		
<i>Podomyrma</i> sp. 3 aff. <i>laevifrons</i> Smith F., 1859	Myrmicinae	<i>Podomyrma</i>	PODO 003	8	7	
<i>Polyrhachis alpheia</i> Smith F., 1863	Formicinae	<i>Polyrhachis</i>	POLY 008	56	3	
<i>Polyrhachis aurita</i> Emery, 1911	Formicinae	<i>Polyrhachis</i>	POLY 047		5	
<i>Polyrhachis debilis</i> Emery, 1887	Formicinae	<i>Polyrhachis</i>	POLY 004	14	26	
<i>Polyrhachis dolomedes</i> Smith F., 1863	Formicinae	<i>Polyrhachis</i>	POLY 020	5	3	
<i>Polyrhachis esuriens</i> Emery, 1897	Formicinae	<i>Polyrhachis</i>	POLY 001	13		
<i>Polyrhachis hostilis</i> Smith, F. 1859	Formicinae	<i>Polyrhachis</i>	POLY 028		1	
<i>Polyrhachis limbata</i> Emery, 1897	Formicinae	<i>Polyrhachis</i>	POLY 031		1	
<i>Polyrhachis lombokensis</i> Emery, 1898	Formicinae	<i>Polyrhachis</i>	POLY 019	8		
<i>Polyrhachis luteogaster</i> Kohout, 2012	Formicinae	<i>Polyrhachis</i>	POLY 010	39		
<i>Polyrhachis mondoi</i> Donisthorpe, 1938	Formicinae	<i>Polyrhachis</i>	POLY 026			18
<i>Polyrhachis mucronata</i> Smith F., 1859	Formicinae	<i>Polyrhachis</i>	POLY 016	3	5	

<i>Polyrbachis nigripes</i> Emery, 1897	Formicinae	<i>Polyrbachis</i>	POLY 039		1
<i>Polyrbachis pallipes</i> Donisthorpe, 1948	Formicinae	<i>Polyrbachis</i>	POLY 021	2	
<i>Polyrbachis queenslandica</i> Emery, 1895	Formicinae	<i>Polyrbachis</i>	POLY 011	8	2
<i>Polyrbachis cf. rossi</i> Donisthorpe, 1948	Formicinae	<i>Polyrbachis</i>	POLY 046		5
<i>Polyrbachis semitestacea</i> Emery, 1900	Formicinae	<i>Polyrbachis</i>	POLY 012		3
<i>Polyrbachis sericata</i> (Guérin-Méneville, 1838)	Formicinae	<i>Polyrbachis</i>	POLY 002	2	19
<i>Polyrbachis waigeuensis</i> Donisthorpe, 1943	Formicinae	<i>Polyrbachis</i>	POLY 015	8	7
<i>Polyrbachis xiphias</i> Smith, F. 1863	Formicinae	<i>Polyrbachis</i>	POLY 029	1	1
<i>Polyrbachis sp. 14 sp. nov.</i> (Myrmatopa, flavicornis group)	Formicinae	<i>Polyrbachis</i>	POLY 014		2
<i>Polyrbachis sp. 23 sp. nov. aff. bubastes</i> Smith F., 1863	Formicinae	<i>Polyrbachis</i>	POLY 023	1	
<i>Polyrbachis sp. 42 cf. sericata</i> (Guérin-Méneville, 1838)	Formicinae	<i>Polyrbachis</i>	POLY 042		4
<i>Polyrbachis sp. 45</i> (Aulacomyrma)	Formicinae	<i>Polyrbachis</i>	POLY 045		2
<i>Polyrbachis sp. 53 aff. menozzii</i> Karavaiev, 1927	Formicinae	<i>Polyrbachis</i>	POLY 053		2
<i>Ponera sp. 1</i>	Ponerinae	<i>Ponera</i>	PONE 001		52
<i>Ponera sp. 2</i>	Ponerinae	<i>Ponera</i>	PONE 002		37
<i>Ponera sp. 3</i>	Ponerinae	<i>Ponera</i>	PONE 003		26
<i>Ponera sp. 4</i>	Ponerinae	<i>Ponera</i>	PONE 004		26
<i>Ponera sp. 7</i>	Ponerinae	<i>Ponera</i>	PONE 007		1
<i>Proceratium austronesicum</i> Baroni Urbani & De Andrade, 2003	Proceratiinae	<i>Proceratium</i>	PROC 001		1
<i>Prolasius sp. 5</i>	Dolichoderinae	<i>Prolasius</i>	PROL 005		4
<i>Pseudolasius breviceps</i> Emery, 1887	Formicinae	<i>Pseudolasius</i>	PSEU 001	1	13
<i>Rhytidoponera strigosa</i> (Emery, 1887)	Ectatomminae	<i>Rhytidoponera</i>	RHYT 001	1	
<i>Rogeria stigmatica</i> Emery, 1897	Myrmicinae	<i>Rogeria</i>	ROGE 001	4	
<i>Solenopsis papuana</i> Emery, 1900	Myrmicinae	<i>Solenopsis</i>	SOLE 004	17	5

<i>Strumigenys cf. racabura</i> Bolton, 2000	Myrmicinae	<i>Strumigenys</i>	STRU 003	2	8	
<i>Strumigenys szalayii</i> Emery, 1897	Myrmicinae	<i>Strumigenys</i>	STRU 002	3	15	
<i>Strumigenys tigris</i>	Myrmicinae	<i>Strumigenys</i>	STRU 007			1
<i>Strumigenys sp. 6 cf. szalayii</i> Emery, 1897	Myrmicinae	<i>Strumigenys</i>	STRU 006			19
<i>Strumigenys sp. 11</i>	Myrmicinae	<i>Strumigenys</i>	STRU 011		1	
<i>Tapinoma cf. indicum</i> Forel, 1895	Dolichoderinae	<i>Tapinoma</i>	TAPI 002	3		
<i>Tapinoma melanocephalum</i> (Fabricius, 1793)	Dolichoderinae	<i>Tapinoma</i>	TAPI 001	2		
<i>Tapinoma sp. 3 aff. williamsi</i> (Wheeler 1935)	Dolichoderinae	<i>Tapinoma</i>	TAPI 003	6	7	
<i>Technomyrmex albicoxis</i> Donisthorpe, 1945	Dolichoderinae	<i>Technomyrmex</i>	TECH 004	1		
<i>Technomyrmex albipes</i> (Smith F., 1861)	Dolichoderinae	<i>Technomyrmex</i>	TECH 002	2		
<i>Technomyrmex cf. cheesmanae</i> Donisthorpe, 1945	Dolichoderinae	<i>Technomyrmex</i>	TECH 008		3	
<i>Technomyrmex difficilis</i> Forel 1892	Dolichoderinae	<i>Technomyrmex</i>	TECH 003	5		
<i>Technomyrmex gilvus</i> Donisthorpe, 1941	Dolichoderinae	<i>Technomyrmex</i>	TECH 005	3		
<i>Technomyrmex mixtus</i> Bolton, 2007	Dolichoderinae	<i>Technomyrmex</i>	TECH 007			55
<i>Tetramorium kydelphon</i> Bolton, 1979	Myrmicinae	<i>Tetramorium</i>	TETR 002	6		
<i>Tetramorium pulchellum</i> Emery, 1897	Myrmicinae	<i>Tetramorium</i>	TETR 012	1	11	
<i>Tetramorium cf. validisculum</i> Emery, 1897	Myrmicinae	<i>Tetramorium</i>	TETR 003		10	
<i>Tetramorium sp. 6 aff. validisculum</i> Emery, 1897	Myrmicinae	<i>Tetramorium</i>	TETR 006	7		
<i>Tetramorium sp. 12 aff. pulchellum</i> Emery, 1897	Myrmicinae	<i>Tetramorium</i>	TETR 016	2		
<i>Tetramorium sp. 20 aff. bicolor</i> Viehmeyer, 1914	Myrmicinae	<i>Tetramorium</i>	TETR 020		1	
<i>Tetramorium sp. 21</i>	Myrmicinae	<i>Tetramorium</i>	TETR 021			1
<i>Tetraoponera atra</i> Donisthorpe, 1949	Pseudomyrmecinae	<i>Tetraoponera</i>	TETP 003	1		
<i>Tetraoponera laeviceps</i> (Smith F., 1859)	Pseudomyrmecinae	<i>Tetraoponera</i>	TETP 001	14	1	
<i>Vollenhovia brachycera</i> Emery, 1897	Myrmicinae	<i>Vollenhovia</i>	VOLL 001	3	2	

<i>Vollenbovia</i> sp. 2	Myrmicinae	<i>Vollenbovia</i>	VOLL 002	4	72
<i>Vollenbovia</i> sp. 3	Myrmicinae	<i>Vollenbovia</i>	VOLL 003		85
<i>Vollenbovia</i> sp. 4	Myrmicinae	<i>Vollenbovia</i>	VOLL 004		4
<i>Vollenbovia</i> sp. 14	Myrmicinae	<i>Vollenbovia</i>	VOLL 014	1	
<i>Vollenbovia</i> sp. 15	Myrmicinae	<i>Vollenbovia</i>	VOLL 015	1	
<i>Vombisidris australis</i> (Wheeler, 1935)	Myrmicinae	<i>Vombisidris</i>	VOMB 004	2	
<i>Vombisidris</i> sp. 2	Myrmicinae	<i>Vombisidris</i>	VOMB 002	2	

**Table A2.** Results of full canonical correspondence analysis (CCA) on the significant explanatory variables (forward-selected at  $p \leq 0.05$ ) of all forest plots combined. The explanatory variables elevation, nest height, tree size and nest types accounted for 7.9% of total variance (6.5% adj.) in ant communities (adjusted p-values used). The significant nest types are marked in italics.

Explanatory factor	% of all variation	% of explained variation	pseudo-F	P (adj.)
Elevation	2.2	26.9	17.2	0.001
Nest height	0.4	5.3	3.5	0.001
Tree size (DBH)	0.3	3.4	2.3	0.002
<i>Carton/ soil/ silk nest on leaves</i>	1.2	14.9	9.7	0.001
<i>Carton/ soil nest on bark</i>	1.1	13.7	8.9	0.001
<i>Under epiphytic roots</i>	0.9	11.0	7.2	0.001
<i>Cavity/ gallery in live twig</i>	0.4	4.3	2.9	0.001
<i>Cavity/ gallery in live branch</i>	0.3	3.1	2.1	0.002
<i>In dead/ rotten twig</i>	0.3	4.0	2.7	0.002
<i>On trunk base</i>	0.3	3.9	2.6	0.045
<i>Under bark</i>	0.3	3.7	2.4	0.001
<i>Cavity/ gallery in trunk</i>	0.2	2.5	1.7	0.003

**Table A3.** Variation partitioning of full dataset across all forest plots to determine the respective contribution of elevation versus the other predictors (DBH, nest height and nest type) from the model in Table A6 to the overall variation in communities (adjusted variability presented).

Explanatory factor	% of all variation (adj.)	% of explained variation (adj.)	Degrees of freedom	Mean Square
Elevation	1.7	26.4	1	0.731
DBH, nest height, nest type	4.4	67.8	11	0.211
Shared variation	0.4	5.8	--	--
Total Explained	6.5	100	12	0.268

**Table A4.** Variation partitioning within each elevation to compare how much tree size (DBH), and nest microhabitats (nest type & nest height) contributed to all variation and explained variation in the different communities. Values were analysed by individual forest plots and mean variability (adjusted) for each elevation are presented.

Elevation	<u>Mean % of all variation</u>			<u>Mean % of explained variation</u>		
	200 m	900 m	1800 m	200 m	900 m	1800 m
DBH	0.3	0.7	2.6	2.2	17.0	16.4
Nest height & nest type	8.7	3.4	12.7	88.0	79.3	81.8
Shared variation	1.0	0.2	0.3	10.0	3.7	1.8
Total explained	9.9	4.3	15.6	100	100	100

**Table A5.** Results of C-Score analysis to examine nesting ant species co-occurrence within trees. Analysis was performed on all trees in a subplot (ALL), and trees of different size classes (small, medium, large). The DBH ranges for small, medium and large trees respectively were as follows: low elevation (5-8 cm, 9-13 cm, 13-100 cm), mid-elevation (5-8 cm, 8.1-14 cm, 14-68 cm), high elevation (5-11 cm, 12-19 cm, 19-96 cm). Two models were run: Fixed model = sim9, Relaxed model = sim2. The fixed model excludes trees with a single species of nesting ants, and the relaxed model includes them. NA marks datasets not tested for C-scores due to low sample size (see methods). Statistically significant at  $P \leq 0.025$  (two-tailed test).

Elevation	Subplot	Tree size	Simulation	Species count	N. trees	Observed C-score	Simulated C-score	SD	SES	Significant
<b>200</b>	<b>200a</b>	<b>Small</b>	<b>sim9</b>	<b>8</b>	<b>10</b>	<b>5.11</b>	<b>4.89</b>	<b>0.10</b>	<b>2.24</b>	<b>TRUE</b>
200	200a	Medium	sim9	13	22	11.97	11.84	0.14	0.97	FALSE
200	200a	Large	sim9	19	36	19.42	19.11	0.27	1.14	FALSE
200	200b	Small	sim9	10	15	6.73	6.79	0.11	-0.49	FALSE
200	200b	Medium	sim9	13	16	5.78	5.76	0.09	0.30	FALSE
<b>200</b>	<b>200b</b>	<b>Large</b>	<b>sim9</b>	<b>27</b>	<b>39</b>	<b>15.17</b>	<b>14.72</b>	<b>0.21</b>	<b>2.19</b>	<b>TRUE</b>
900	900a	Small	sim9	22	30	9.24	9.27	0.09	-0.42	FALSE
900	900a	Medium	sim9	26	44	17.70	17.67	0.10	0.27	FALSE
900	900a	Large	sim9	56	69	24.21	24.19	0.14	0.11	FALSE
1800	1800a	Small	sim9	3	3	NA	NA	NA	NA	NA
1800	1800a	Medium	sim9	7	13	11.00	10.98	0.23	0.11	FALSE
1800	1800a	Large	sim9	15	37	29.90	30.04	0.39	-0.37	FALSE
1800	1800c	Small	sim9	3	5	NA	NA	NA	NA	NA
1800	1800c	Medium	sim9	8	10	5.11	5.06	0.11	0.39	FALSE
1800	1800c	Large	sim9	9	17	13.17	13.19	0.17	-0.15	FALSE
200	200a	ALL	sim9	35	86	33.35	32.85	0.27	1.85	FALSE
200	200b	ALL	sim9	39	83	29.38	29.08	0.22	1.33	FALSE
<b>900</b>	<b>900a</b>	<b>ALL</b>	<b>sim9</b>	<b>70</b>	<b>157</b>	<b>53.24</b>	<b>52.46</b>	<b>0.27</b>	<b>2.95</b>	<b>TRUE</b>
1800	1800a	ALL	sim9	15	59	72.47	72.36	0.67	0.15	FALSE
1800	1800c	ALL	sim9	10	34	41.96	41.41	0.33	1.62	FALSE
<b>200</b>	<b>200a</b>	<b>Small</b>	<b>sim2</b>	<b>15</b>	<b>47</b>	<b>15.20</b>	<b>13.60</b>	<b>0.51</b>	<b>3.16</b>	<b>TRUE</b>
<b>200</b>	<b>200a</b>	<b>Medium</b>	<b>sim2</b>	<b>14</b>	<b>38</b>	<b>19.53</b>	<b>18.05</b>	<b>0.73</b>	<b>2.01</b>	<b>TRUE</b>
200	200a	Large	sim2	22	54	21.82	21.32	0.73	0.67	FALSE
<b>200</b>	<b>200b</b>	<b>Small</b>	<b>sim2</b>	<b>18</b>	<b>71</b>	<b>23.74</b>	<b>20.69</b>	<b>0.61</b>	<b>5.02</b>	<b>TRUE</b>
<b>200</b>	<b>200b</b>	<b>Medium</b>	<b>sim2</b>	<b>19</b>	<b>39</b>	<b>9.84</b>	<b>9.13</b>	<b>0.32</b>	<b>2.23</b>	<b>TRUE</b>
200	200b	Large	sim2	28	56	18.54	18.59	0.49	-0.10	FALSE
<b>900</b>	<b>900a</b>	<b>Small</b>	<b>sim2</b>	<b>26</b>	<b>67</b>	<b>16.55</b>	<b>15.52</b>	<b>0.39</b>	<b>2.62</b>	<b>TRUE</b>
900	900a	Medium	sim2	29	71	22.87	22.15	0.41	1.77	FALSE
<b>900</b>	<b>900a</b>	<b>Large</b>	<b>sim2</b>	<b>56</b>	<b>74</b>	<b>25.03</b>	<b>26.31</b>	<b>0.25</b>	<b>-5.20</b>	<b>TRUE</b>
<b>1800</b>	<b>1800a</b>	<b>Small</b>	<b>sim2</b>	<b>9</b>	<b>36</b>	<b>19.92</b>	<b>16.01</b>	<b>0.95</b>	<b>4.11</b>	<b>TRUE</b>
1800	1800a	Medium	sim2	9	26	18.31	17.37	1.26	0.74	FALSE
1800	1800a	Large	sim2	15	44	35.51	35.50	1.50	0.01	FALSE
<b>1800</b>	<b>1800c</b>	<b>Small</b>	<b>sim2</b>	<b>6</b>	<b>40</b>	<b>38.33</b>	<b>24.46</b>	<b>3.55</b>	<b>3.89</b>	<b>TRUE</b>
<b>1800</b>	<b>1800c</b>	<b>Medium</b>	<b>sim2</b>	<b>10</b>	<b>40</b>	<b>23.16</b>	<b>19.43</b>	<b>1.11</b>	<b>3.35</b>	<b>TRUE</b>
<b>1800</b>	<b>1800c</b>	<b>Large</b>	<b>sim2</b>	<b>9</b>	<b>29</b>	<b>24.39</b>	<b>21.79</b>	<b>1.44</b>	<b>1.80</b>	<b>TRUE</b>



**Table A6.** ANOVA to select best GLM model testing the effect of elevation on species dissimilarity between trees (Sørensen dissimilarity index). Model 1 was a significantly better fit than the null model. Model 0: dissimilarity ~ + 1, Model 1: dissimilarity ~ Elevation

Model	Residual df	Residual deviance	df	Deviance	P value
0	63406	25119			
1	63404	24415	2	704.05	<0.001

**Table A7.** Results of above GLM model 1 testing effect of elevation on species dissimilarity between trees. (Sørensen dissimilarity index).

	Estimate	Standard Error
200 m (intercept)	2.34	0.015
900 m	0.14	0.022
1800 m	-0.73	0.023

**Table A8.** General linear mixed models (glmmTMB) testing the effect of tree size and elevation on species richness per tree. Models compared with anova (mod<sub>a</sub>, mod<sub>b</sub>, test=chisq).

Model	Independent variable(s)	DF	AIC	Deviance	Test against	Chisq	Chi DF	p
Mod.0	+1	3	3868.8	3862.8	-	-	-	-
Mod.1	DBH	4	3531.7	3523.7	Mod.0	339.15	1	<0.01***
Mod.2	Elevation	5	3859.9	3849.9	Mod.0	12.927	2	<0.01**
Mod.3	A + B	6	3520.9	3508.9	Mod.1	14.836	2	<0.01***
Mod.4	A+B+A:B	8	3555.6	3498.6	Mod.3	10.307	2	<0.01***

**Table A9.** General linear mixed models (glmmTMB) testing the effect of tree size and elevation on nest abundance. Models compared with anova (mod<sub>a</sub>, mod<sub>b</sub>, test=chisq).

Model	Independent variable(s)	DF	AIC	Deviance	Test against	Chisq	Chi DF	p
Mod.0	+1	3	4366.5	4360.5	-	-	-	-
Mod.1	DBH	4	4000.2	3992.2	Mod.0	368.32	1	<0.01***
Mod.2	Elevation	5	4357.8	4347.8	Mod.0	12.716	2	<0.01***
Mod.3	A + B	6	3990.3	3978.3	Mod.1	13.867	2	<0.01***
Mod.4	A+B+A:B	8	3990.4	3974.4	Mod.3	3.8692	2	0.1445

**Table A10.** General linear mixed models (glmmTMB) testing the effect of tree size and elevation on nest diversity. Models compared with anova (mod<sub>a</sub>, mod<sub>b</sub>, test=chisq).

Model	Independent variable(s)	DF	AIC	Deviance	Test against	Chisq	Chi DF	p
Mod.0	+1	3	3315.5	3309.5	-	-	-	-
Mod.1	DBH	4	3100.8	3092.8	Mod.0	216.66	1	<0.01***
Mod.2	Elevation	5	3306.0	3296.0	Mod.0	13.44	2	<0.01**
Mod.3	A + B	6	3086.2	3074.2	Mod.1	18.583	2	<0.01***
Mod.4	A+B+A:B	8	3080.7	3064.7	Mod.3	9.5266	2	<0.01**